



IP-One

A new dimension in Gαq signaling

What every GPCR
scientist should know
about IP-One

Cisbio was the first to enter and develop the market for TR-FRET and inspire scientists with HTRF possibilities. Launched in 1990, HTRF is now a leading tool for screening, target ID and lead validation, and Cisbio provide global researchers with a broad range of assays and depth in scientific expertise.

For the last two decades, Cisbio has delivered unparalleled expertise in screening and continues to market innovative technologies. Cisbio supports the best minds in screening and provides relevant, easy to use kits for all major therapeutic areas.

Today, as the premier TR-FRET technology on the market, HTRF has been applied to a variety of application including GPCRs, kinases, epigenetics, biotherapeutics and quantification of a range of biomarkers, including cytokines. Researcher now rely on Cisbio assays to deliver rapid and accurate results with high sensitivity in an easy-to-use format that delivers robust and accurate results.

Kits
198

Reagents
380

References
1650

A new dimension in Gαq signaling

The diagram illustrates a complex signaling pathway. At the top, a cell membrane is shown with various receptors and proteins. A yellow wavy line represents a G-protein (Gq) that activates PLC. PLC converts PIP2 into IP3 and Diacylglycerol (DAG). IP3 binds to the ER, releasing Ca²⁺. IP1 is formed from IP3 and DAG, and is inhibited by LiCl. IP1 is converted to Myo-inositol and IP2. IP2 is converted to P-ERK. P-ERK is also formed from P-Akt. P-Akt is formed from B-arrestin. B-arrestin is formed from the receptor. The receptor is also inhibited by MP (Methamphetamine) and IBMX (Isobutylmethylxanthine). The pathway is regulated by ATP and AMP. The diagram shows the following components and interactions:

- Receptor**: A grey wavy line on the membrane.
- Gq**: A yellow wavy line representing a G-protein.
- PLC**: A blue oval protein.
- PIP2**: A yellow oval lipid.
- IP3**: A yellow oval lipid.
- IP1**: A blue circle lipid.
- IP2**: A yellow oval lipid.
- Myo-inositol**: A yellow oval lipid.
- P-ERK**: A blue circle protein.
- P-Akt**: A blue circle protein.
- B-arrestin**: A grey oval protein.
- ER**: A grey oval protein.
- Ca²⁺**: Grey dots representing calcium ions.
- MP**: A grey oval protein.
- IBMX**: A grey oval protein.
- ATP**: A grey oval protein.
- AMP**: A grey oval protein.
- LiCl**: A blue circle protein.

Interactions:

- Receptor → B-arrestin → P-Akt → P-ERK
- Gq → PLC → PIP2 → IP3 → IP1 → IP2 → P-ERK
- IP1 → Myo-inositol
- IP1 → IP2
- IP2 → P-ERK
- P-Akt → P-ERK
- MP → Receptor
- IBMX → Receptor
- ATP → Adenylate cyclase → cAMP → PKA → P-ERK
- AMP → Adenylate cyclase
- LiCl → IP1



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Overview

Cisbio offers an innovative approach with IP-One, a non-radioactive, homogeneous, inositol-phosphate accumulation assay for Gq signaling. This kit has been shown to offer superior benefits over traditional assays, making it the ideal tool for your research.

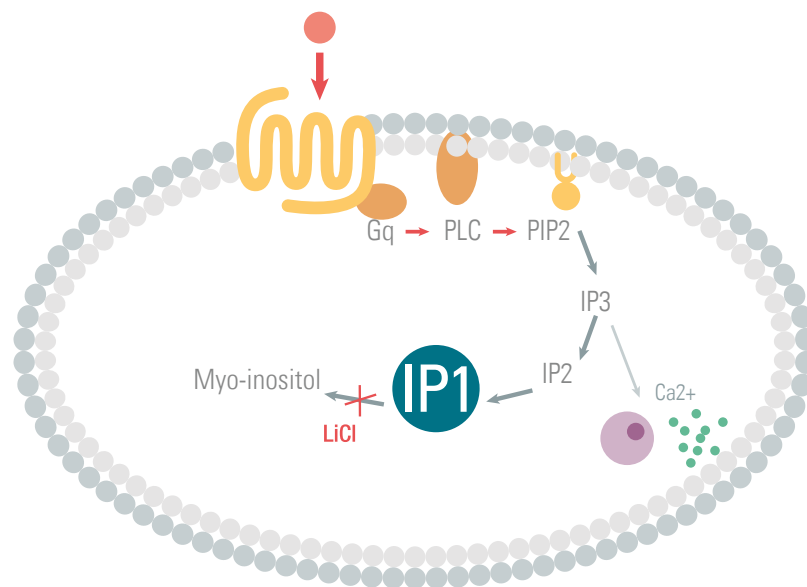




Signaling pathway

Cisbio's IP-One is a reliable and recognized tool for **investigating the Gq/11 coupled receptor**.

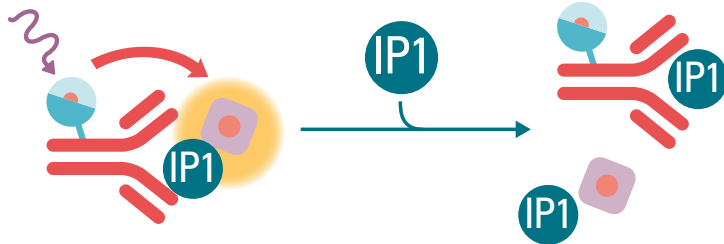
Cisbio has developed a **GPCR platform**, based on the company's HTRF® technology, to measure second messenger accumulation. This platform includes IP-One, a very unique, non-isotopic assay. This bioassay kit specifically measures IP1 levels in cells. Over the past decade, IP-One has demonstrated its efficacy on numerous occasions for discovering and/or characterizing new compounds that target GPCRs.





Assay Format

IP-One is a patented **competitive immunoassay** involving a specific antibody terbium cryptate (donor) and IP1 coupled to d2 (acceptor).



In a standard state, the interaction between the donor and the acceptor produces a fluorescent signal. When unlabeled IP1 is present in the sample (due to GPCR activation), binding competitively to the antibody and taking the place of the labeled IP1. Fluorescence then decreases, as the donor is no longer in close proximity to the acceptor. The range of the decrease is directly related to IP1 accumulation.



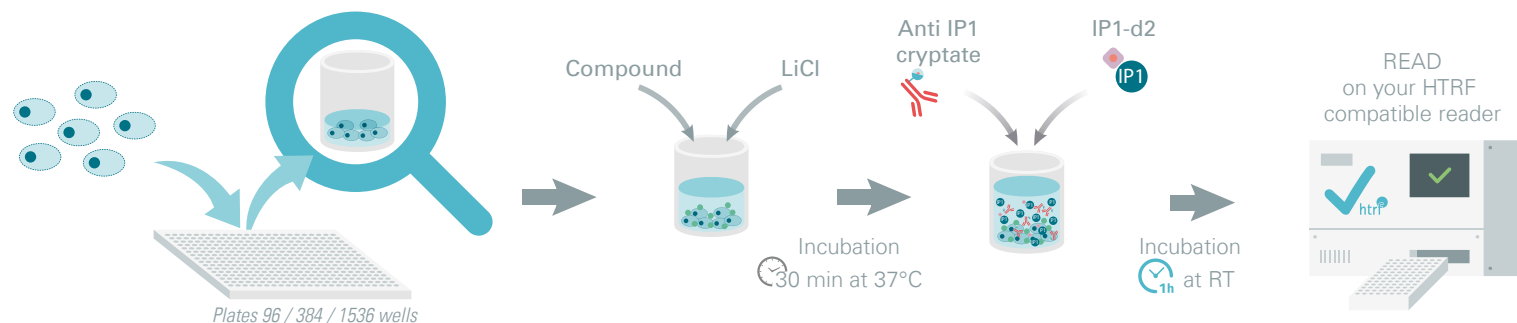


Assay Protocol

As with most HTRF-based assays, IP-One is very easy to set up. The IP-One assay protocol involves two incubation steps:

- Cell stimulation by the ligand or target compounds
- IP1 detection using HTRF reagents

The kit provides everything you need, including the Lithium Chloride used to prevent IP1 degradation.



This protocol requires **only a single, one-hour incubation period** following cell stimulation.

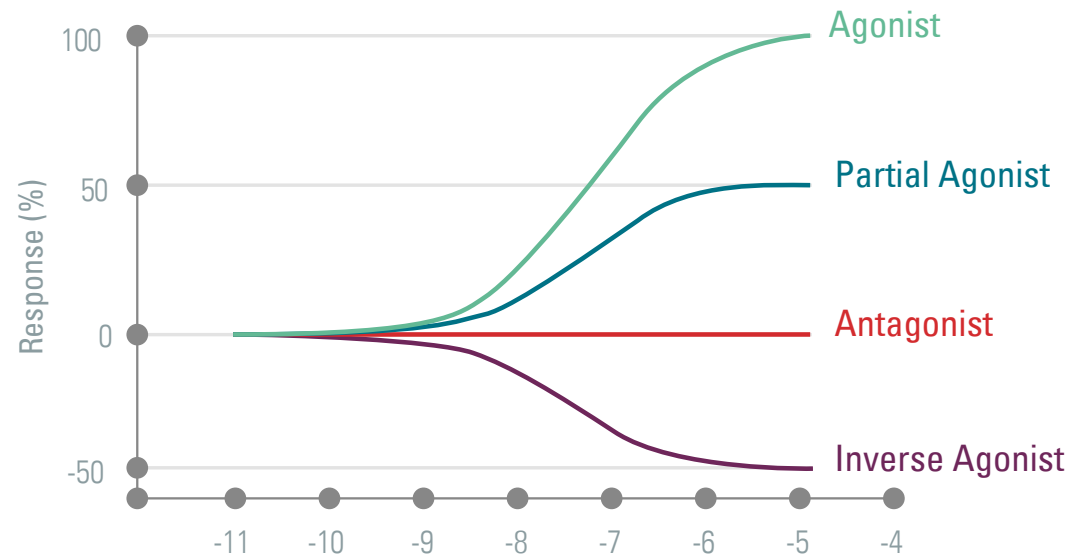




IP-One enables investigation of all classes of pharmaceutical compounds

IP-One makes it possible to assess all classes of pharmaceutical compounds, namely:

- Agonists
- Antagonists
- Allosteric modulators
- Inverse agonists
- Slow-acting binders





Benefits

Choosing IP-One for investigating Gq/11 coupled receptors offers **numerous benefits**:

- Simple protocol and assay kit
- Miniaturization down to 1536-well plates
- Receptor proximal marker
- Compatible with primary and non-recombinant cell lines
- And more...



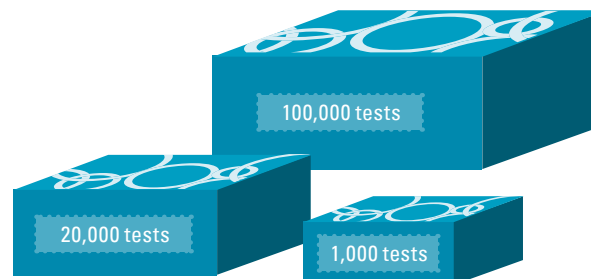
IP-One brings you closer **to discovering drugs**.



Packaging

To better answer all your needs, IP-One is available in different packaging, from 1,000 tests to 100,000 tests.

We are also able to deliver the exact volume of reagent, in a single batch, needed to match your screening requirements.



Packaging	Part#
IP-One - Gq kit - 1,000 tests	62IPAPEB
IP-One - Gq kit - 20,000 tests	62IPAPEC
IP-One - Gq kit - 100,000 tests	62IPAPEJ

There will always be a **solution**
adapted to your needs.





High Throughput Screening

When imagining an assay that is HTS compatible, the first thing that most people think about is performance and miniaturization. Of course, IP-One answers these needs, but there is more. Reagent stability and signal stability may seem less critical at first glance, but they are both highly necessary for successful screening.

This section provides an in-depth focus on the points that make IP-One a true, trusted assay for HTS.





Ease-of-use

IP-One: a homogeneous assay

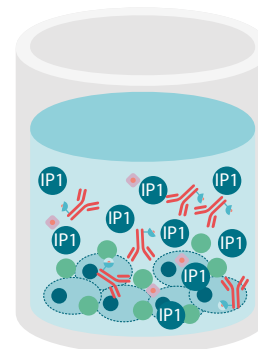
As mentioned earlier, IP-One is a very simple assay to use. Some suppliers' assays may be limited with respect to HTS requirements, but not the HTRF assays on which IP-One is based!

Homogeneous “**add-and-read**” assays are well-suited for HTS, as they avoid filtration, separation, and wash steps that can be both time-consuming and difficult to automate.

Using IP-One for your screen offers several **advantages**:

- No complex robotic plate washers required
- Eliminates equipment maintenance costs
- Saves you time by minimizing the number of steps needed to run the assay

ADD and
READ

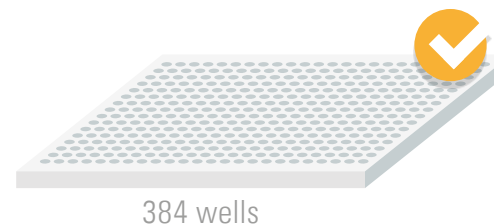
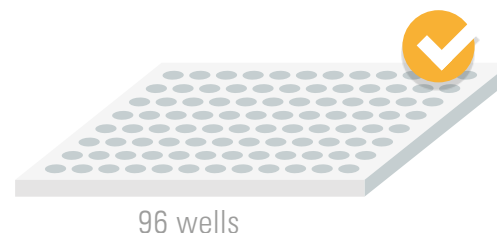


Miniaturization

Miniaturization lowers reagent consumption and increases throughput. Cisbio considers that an assay must **adapt to your needs**, not the other way around!

IP-One can be implemented from 96w (100 μ L or 20 μ L using our new HTRF 96-well low volume plate) to 384w or even 1536w plates (10 μ L), depending on your specific situation.

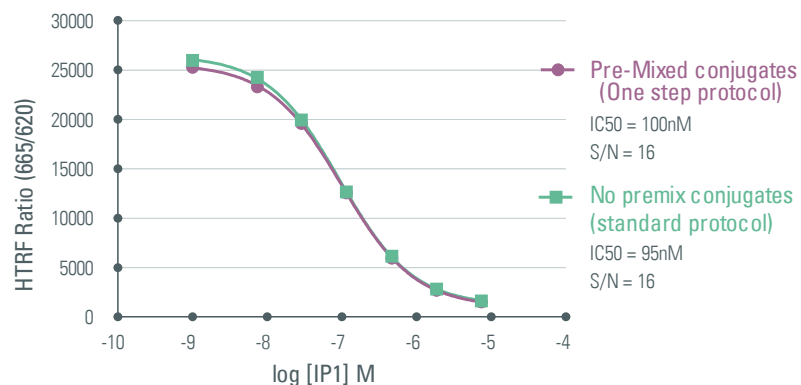
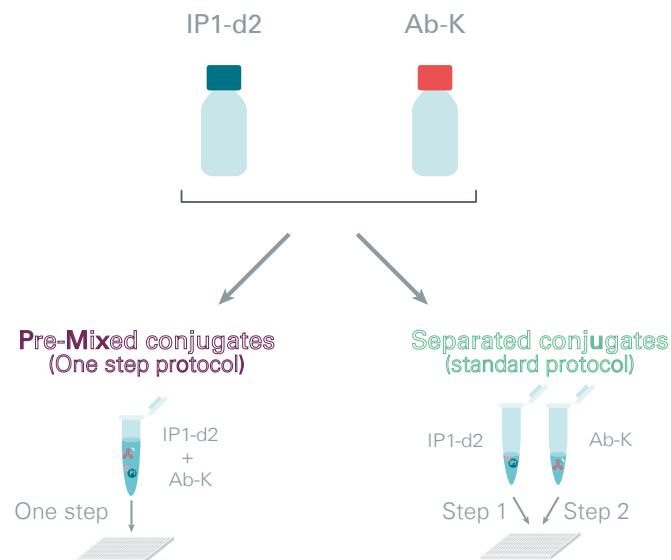
Cisbio always has **the right solution** for you.



IP-One can make it even easier

The classic protocol for IP-One requires preparation of two working solutions (one for IP1-d2, the other for Ab-K), followed by two successive dispensing operations.

Here, we demonstrate the possibility of premixing your conjugates, which makes it easier to use IP-One, and streamlining the process without compromising your results.



Get the full details!

Download & read the technical note at
[cisbio.com](https://www.cisbio.com)

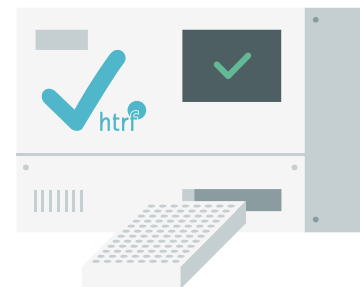
IP-One: no expensive plate reader required

While an intracellular calcium assay requires the use of a costly kinetic fluorescence assay, the IP-one assay can be performed using a **standard fluorescence plate reader**.

In order to broaden HTRF readout capabilities for use in life sciences, Cisbio has established partnerships with several detection system manufacturers and meticulously certified many instruments for HTRF readout, from dedicated readers to multimode systems.



is **compatible** with more than
30 different readers
from **7** different brands



+ visit our website to learn more





Performance

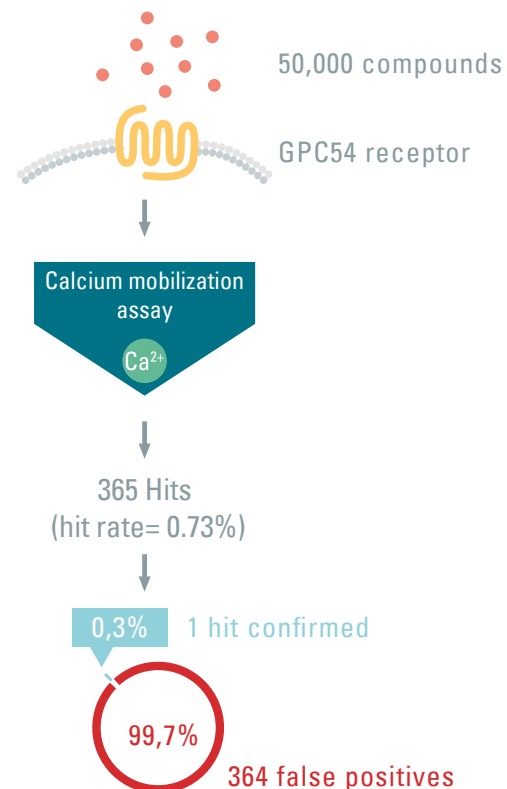
Fewer false positives:

IP-One brings more specificity to your screens

There is no need to demonstrate how false positives can have a negative effect on your screen: they lead to extra work and incur additional cost, as the false positives need to be eliminated.

IP-One simply delivers fewer false positives than calcium flux technologies, practically eliminating the need to counter-screen the hits obtained from primary screening.

In this example, representative of a typical workflow with a calcium flux screen, 50,000 compounds were screened and 365 hits were obtained. In reality, 99.7% of the hits were false positives. Only one hit was confirmed with multiple technologies (including IP-One).



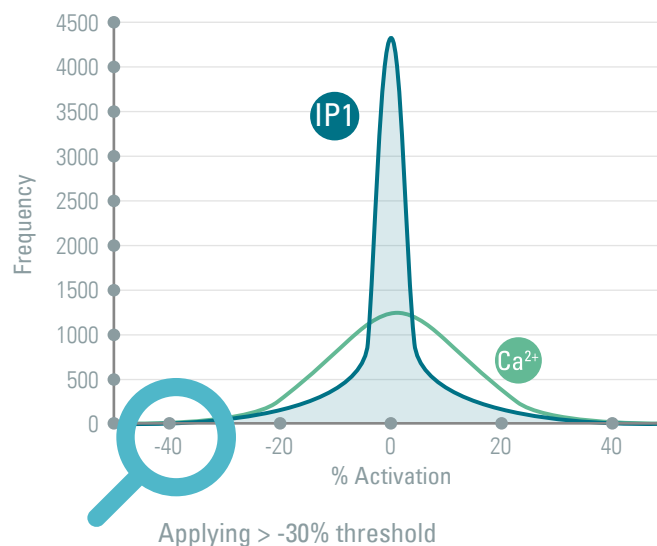
This workflow was adapted from: K J. Cassutt et al «Identifying Nonselective Hits from a Homogeneous Calcium Assay Screen,»
Journal of Biomolecular Screening 12(2) (2007)



The high false positive rate associated with calcium flux technologies leads to an erroneous assessment of the discovery rate, resulting in an inflated distribution of hits.

IP-One reduces the false positive rate, displaying a lower hit rate but with almost only specific hits.

Analysis of 7744 compounds from Napr collection in IP1 and Calcium measurement



IP1 *Hit rate* 0,34%

- Lower hit rate
- Specific hits

Ca²⁺ *Hit rate* 4,58%

- Higher hit rate
- False positive
- Compounds interferences

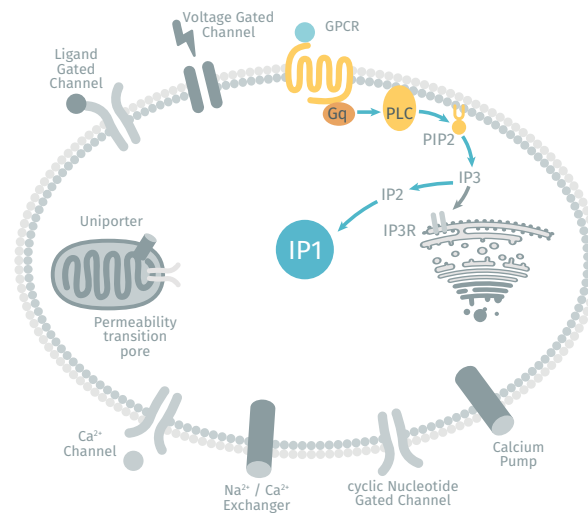
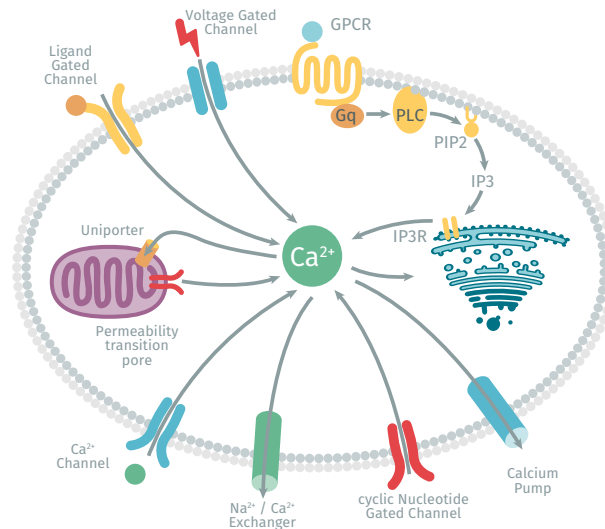


Numerous of reasons can be found to explain the high rate of false positives obtained with calcium flux assays. Calcium responses are complex in nature. Calcium release is frequently triggered by non-G protein mechanisms (Ca^{2+} permeable channels, Ca^{2+} pumps, and Ca^{2+} transporters). Compounds engaging these receptors are said to be “off-target”.

IP-One **ensures signal specificity** reducing the high rate of false positives.

It only measures IP1, produced solely by phospholipase enzyme (PLC) action and mediated by GPCRs.

Using calcium flux technology, Ca^{2+} ionophores and compounds that permeabilize the cellular membrane appear as false positives in agonist screening assays. IP-One ensures that these compounds remain undetected.

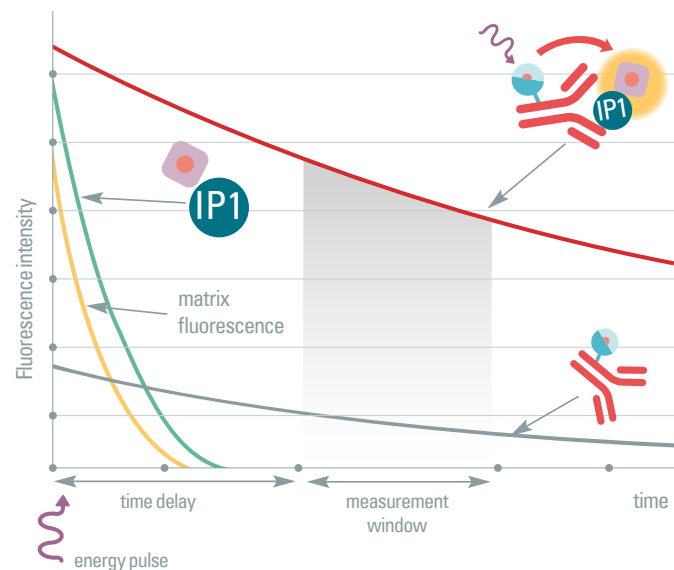


Using calcium flux assays, you may be confronted with:

- Blocked tips that lead to false positives in inhibitor screens, or false negatives in agonist screens.
- Auto-fluorescent compounds that appear as false positives in agonist mode.

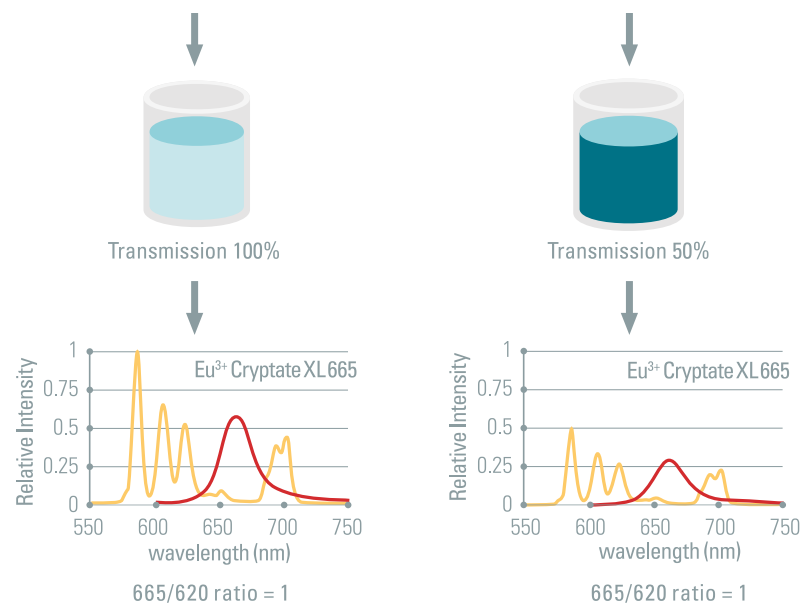
Standard compound libraries used for a screen may contain a significant number of auto-fluorescent compounds. Based on a time-resolved signal, IP-One eliminates the emission signal from most auto-fluorescent compounds automatically, thus lowering the rate of false positives.

The energy pulse from the excitation source (flash lamp or laser) is immediately followed by a time delay, allowing the decay of interfering short-lived fluorescence, such as from compounds, proteins, or medium.



Another advantage of HTRF is that it uses a specific ratio, which protects results from any inner filter effects. HTRF emissions are measured at two wavelengths: **620nm (donor)** and **665nm (acceptor)**.

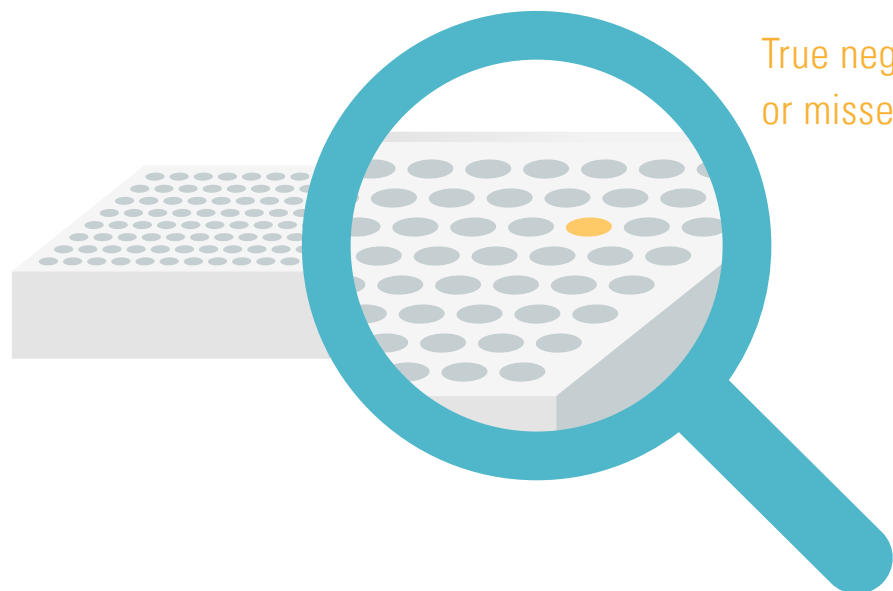
Dual-wavelength measurements are extremely advantageous and help compensate for well-to-well variations. Filter effects may occur when highly colored compounds are used, leading to a decrease in the excitation energy delivered. The fluorescence ratio **compensates for this phenomenon** because both acceptor and donor emissions are affected in the same manner.



Low false negative rate: don't miss your hits anymore

Having a high false positive rate may cost you time and money, but your hit is still there in the end. On the other hand, a high false negative rate can have disastrous impact on your screen. What if the molecule you were looking for was really there, your future block-buster, but you missed it because it was undetected by your screening technology?

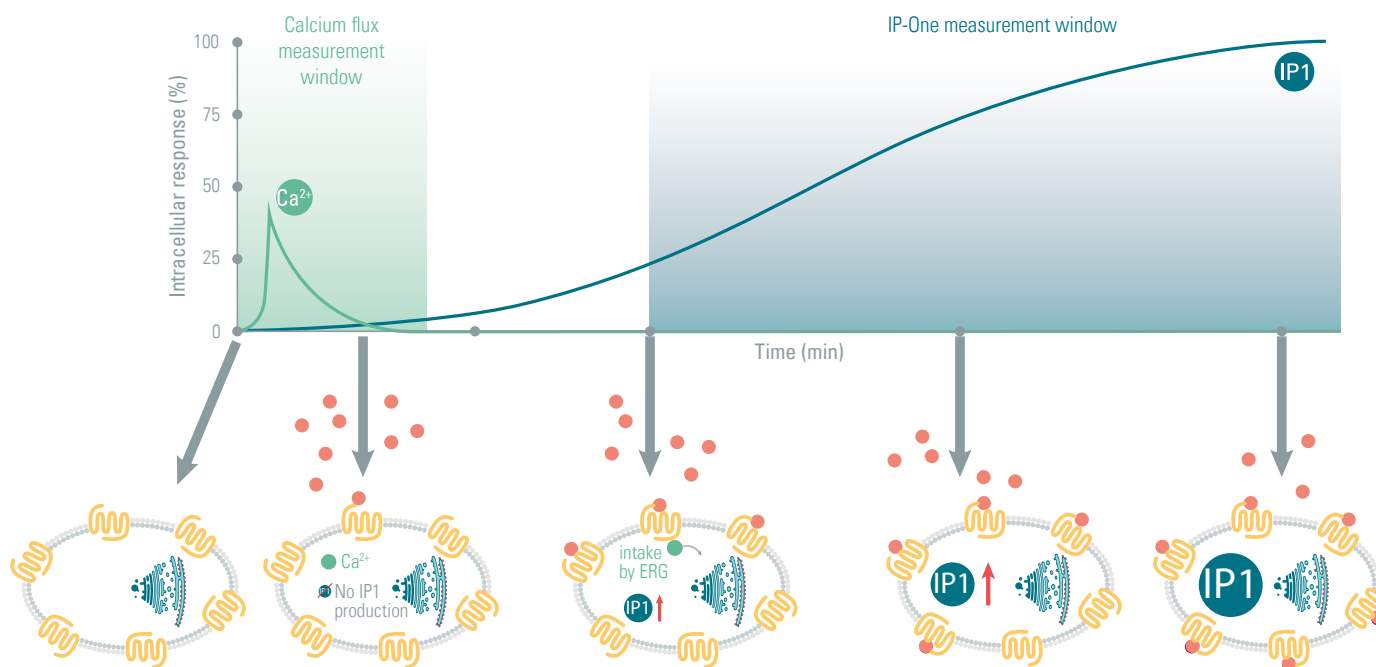
The consequences of **false negatives** are **too serious** to tolerate.



True negative
or missed hit?

Interpretations of calcium assay results may be hindered because of the non-equilibrium state. The absence of equilibration results in biased IC50 for inhibitors, and false negatives in screening campaigns in which important molecules may be lost forever.

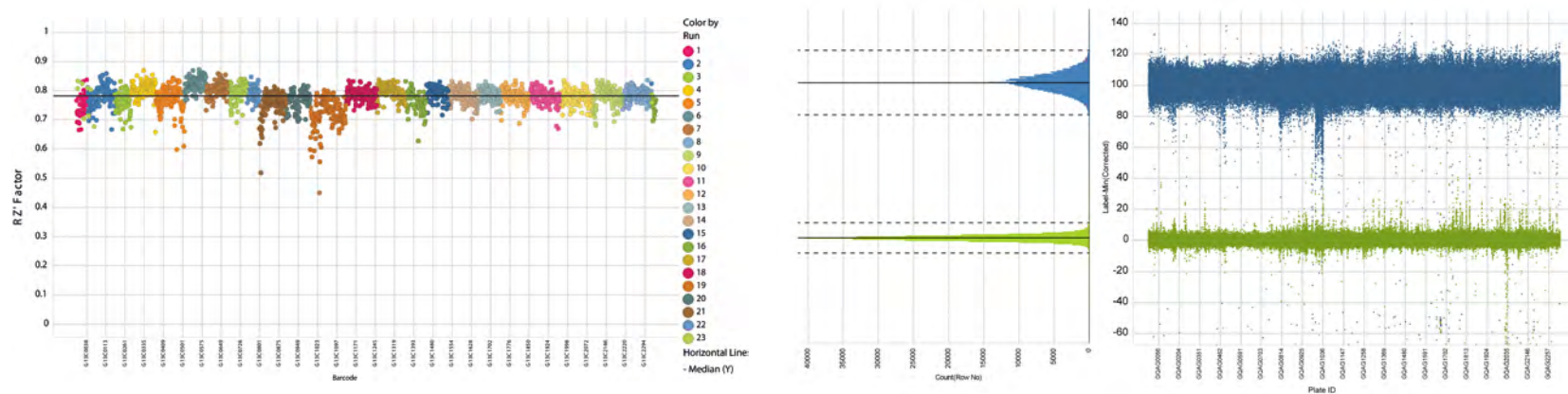
IP-One is **built on an accumulation assay** and allows you to record every possible hit, including slow-acting compounds with **true pharmacology** associated with **accurate IC50 determination**.



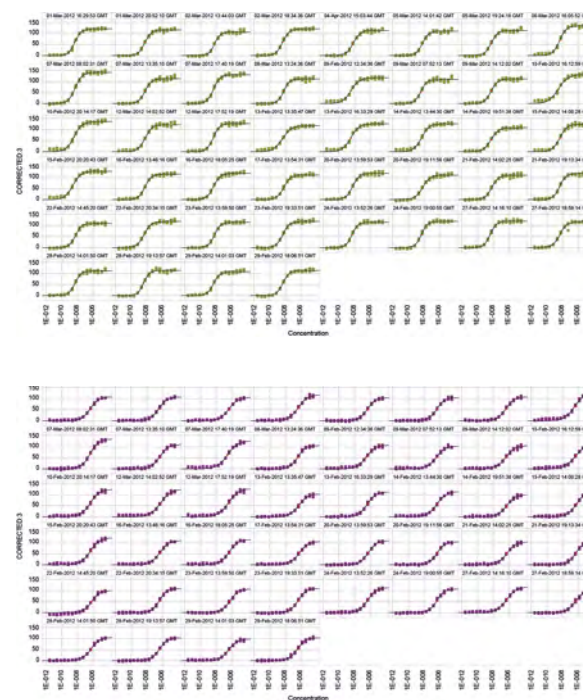
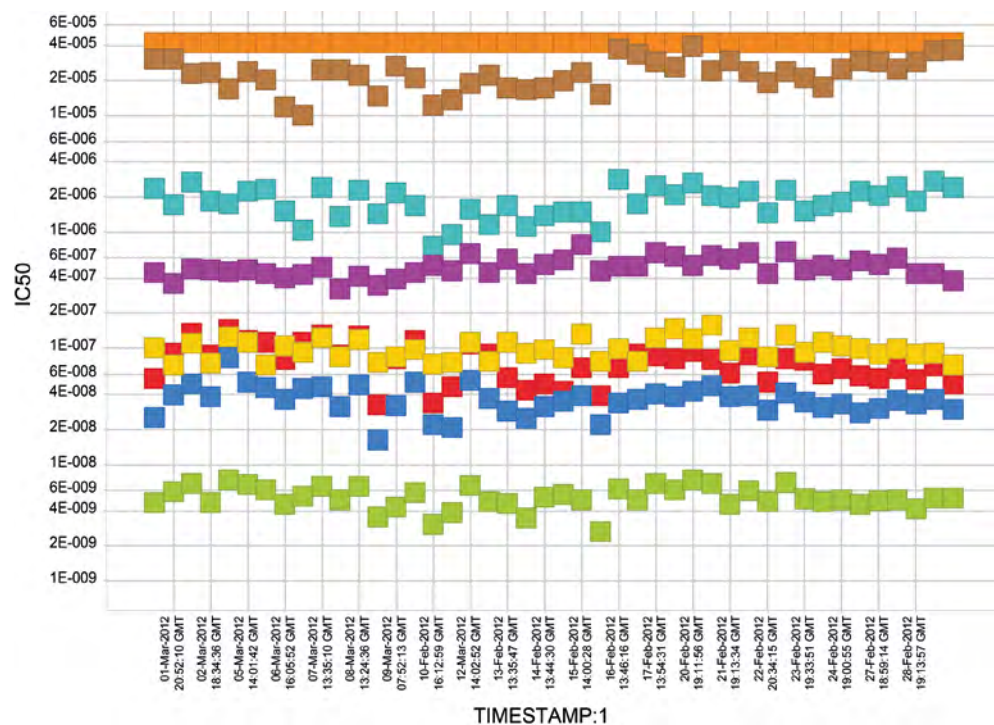
Robustness: IP-One delivers high quality Z' and high reproducibility

The Z' score is perhaps the most widely-used statistical parameter for measuring assay robustness for HTS. In one case study by Bayer, IP-One screen proved excellent assay quality providing high robust Z' factors in every run (~ 0.78) and broad separation of controls.

Here are some examples of data scatters obtained by Bayer in one case study:



Bayer also obtained constant IC50 values for reference compounds over the entire screening in one case.



Robustness: IP-One Z' versus calcium assay Z'

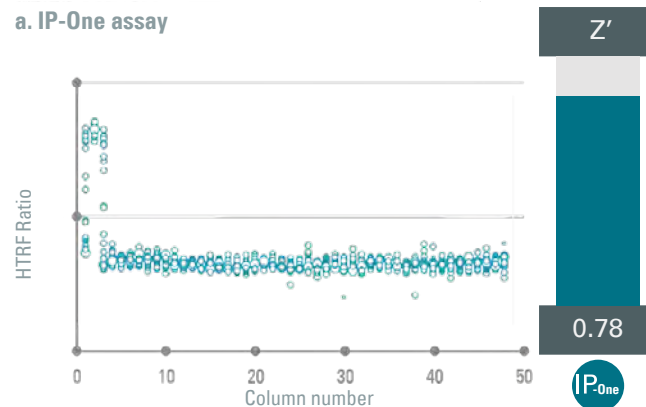
By measuring Z' factor reliably, **IP-One stands out as a very robust assay.**

Several reasons can explain the differences in robustness between these two assays. For example, conventional calcium mobilization protocols are multi-step procedures, and each additional step carries the risk of increased assay variability.

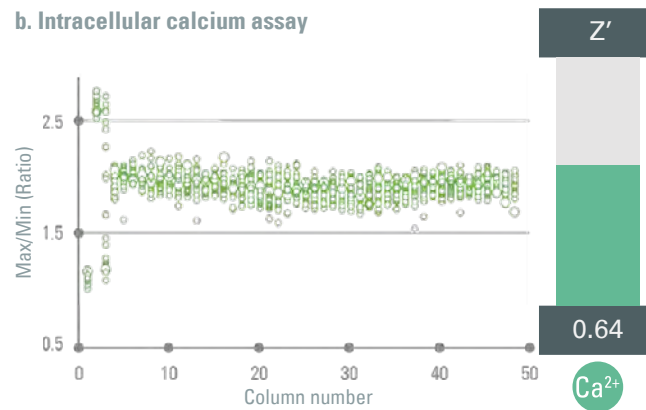
This inevitably decreases Z':

- **Removal of cells** from wells during the wash procedure to purge excess dye
- **Reduced responsiveness** (competence) of cells after washing due to perturbation
- **Spontaneous calcium flux** upon the addition of buffer or compound
- **Incomplete washing** resulting in a significant signal drop upon the addition of test compound
- **Asymmetric release of probenecid** by cells

a. IP-One assay



b. Intracellular calcium assay



Adapted from: Liu K (2008), Curr Chem Genomics. Comparison on Functional Assays for Gq-Coupled GPCRs by Measuring Inositol Monophosphate-1 and Intracellular Calcium in 1536-Well Plate Format.



Robustness: Literature

A significant body of literature has demonstrated how **IP-One robustness outperforms calcium flux technologies**.

Some significant examples are listed below.

- *Bergsdorf* «A One-Day, Dispense-Only IP-One HTRF Assay for High-Throughput Screening of Gq Protein-Coupled Receptors: Towards Cells as Reagents.», *Assay Drug Dev Technol* (2008).
- *Liu K* «Comparison on Functional Assays for Gq-Coupled GPCRs by Measuring Inositol Monophosphate-1 and Intracellular Calcium in 1536-Well Plate Format.», *Current Chemical Genomics* (2008).
- *Cassutt* «Identifying Nonselective Hits from a Homogeneous Calcium Assay Screen.», *Journal of Biomolecular Screening* (2007).
- *Kuohung* «A high-throughput small-molecule ligand screen targeted to agonists and antagonists of the G-protein-coupled receptor GPR54.», *Journal of Biomolecular Screening* (2010).
- *Thomsen* «Functional assays for screening GPCR targets.», *Current Opinion in Biotechnology* (2005).



Stability: Cisbio guarantees its reagent stability

When you screen thousands of compounds, it may take several hours before the reagents are added to all of the assay plates. Cisbio reagent stability during and beyond daily screening operations (i.e. 8 hour shifts) has been confirmed.

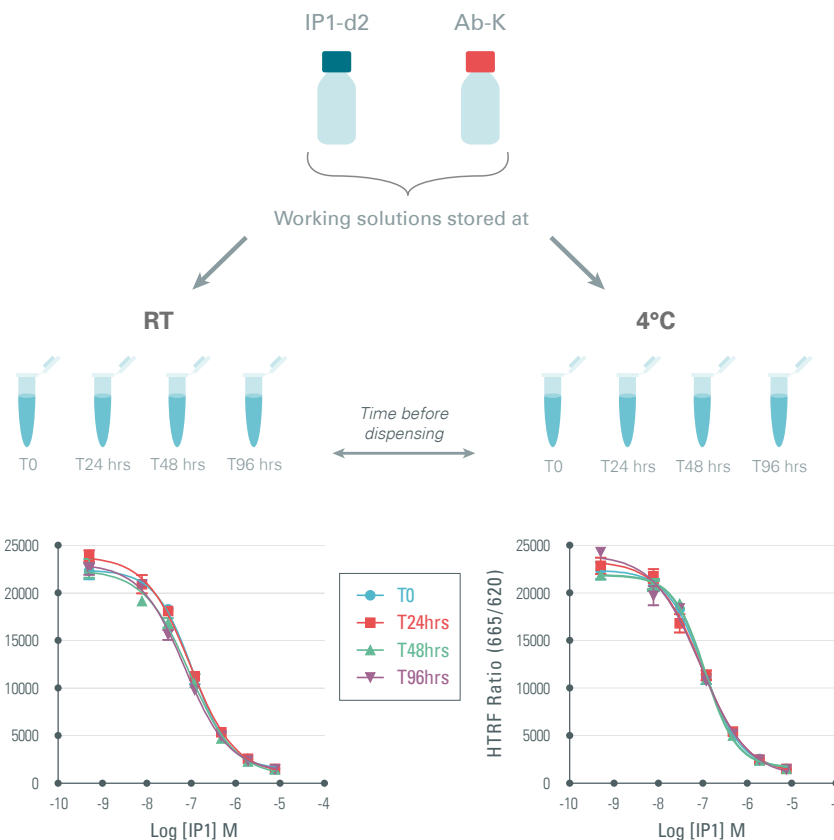
Reflecting a typical screening campaign, the following study describes a series of stability tests conducted to confirm that IP-One reagents are stable:

- IP-One reagents are stable for several days at 4°C and room temperature
- IP-One delivers consistent performance over time



Get the full details!

Download & read the technical note at
cisbio.com



Stability: Signal stability is key for a successful HTS campaign

An unstable signal means that plates must be read shortly after adding reagent, potentially decreasing throughput. Additionally, in the event of a problem on the screening chain, the signal may be lost forever.

Plates were incubated at room temperature for various lengths of time (from 1 hr. to 8 days) before being read on an HTRF compatible reader. The signal, as well as the IP1 standard curve assay window, remain stable over time, demonstrating the same performance.

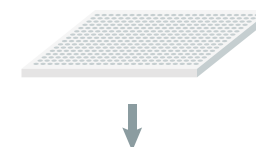
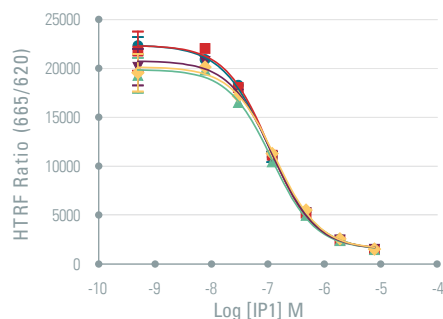


Plate incubated at RT
and read after:

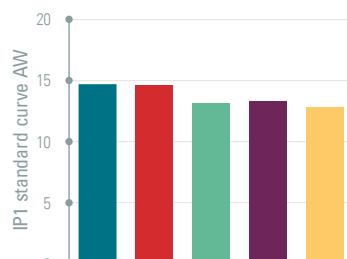
- 1 h
- 1 day
- 2 days
- 5 days
- 8 days



IP-One assay signal stays stable
for days after reagent addition



IP-One assay window stays stable
for days after reagent addition



- With its stable signal, IP-One protects against unforeseen issues that may occur on the screening platform
- IP-One is highly flexible and allows you to read your results at any point in time





Reagent availability

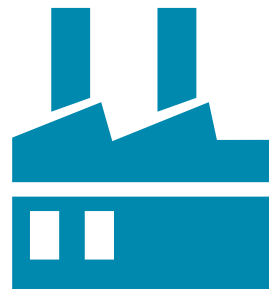
Our kits are **always in stock**

All IP-One reagents, including the anti-inositol-1 antibody, are produced by Cisbio laboratories directly, which means that we can guarantee **reagent accessibility** and **stock availability**.

Sourcing multiple batches of reagents affects HTS performance and reproducibility. Cisbio understands that having access to a single batch for HTS is a requirement.

Cisbio delivers the exact volume of reagent, in a single batch, needed to match your screening requirements.

We own
OUR
antibody





Reagent quality

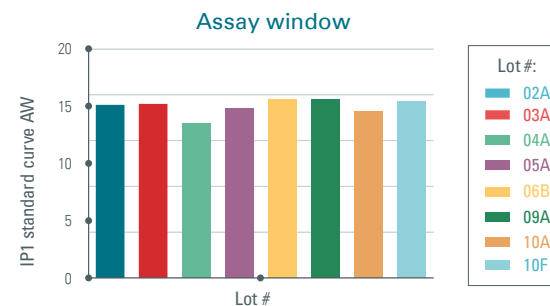
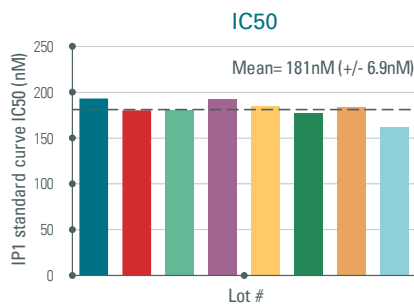
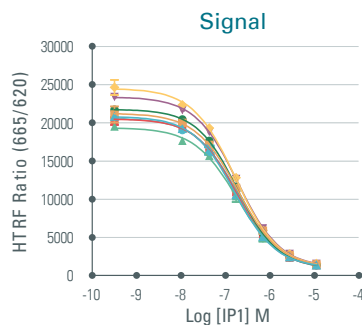
Batch-to-batch quality control

We invest significant resources to guarantee the quality of our screening reagents. All Cisbio reagents are required to pass **strict quality metrics**, with every batch tracked for:

- Labeling efficiency
- Assay signal
- IC50 accuracy
- Batch-to-batch reproducibility



IP1 standard curves (Batch to batch reproducibility):





Webinar

High-Throughput Screening of GPCRs

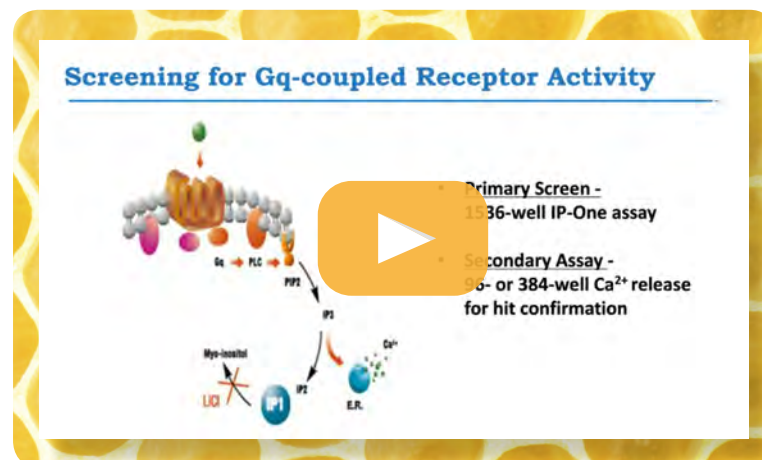
A webinar discussing High-Throughput Screening of GPCRs featuring:

- **Sylvie Jezequel-Sur** (MS, Associate Principal Scientist - Merck & Co),
- **R. Jason Kirby** (Senior Associate Scientist - Sanford Burnham Prebys Medical Discovery Institute),
- **Dr Edward Ainscow** (Director - Genomics Institute of the Novartis Research Foundation)

is available to watch on demand.



Watch the full webinar
at cisbio.com





Therapeutic area

Needs differ with the target field. At Cisbio, we know the importance of making assays that adapt to your needs, not the opposite.

IP-One has numerous advantages that will help you in your daily research.



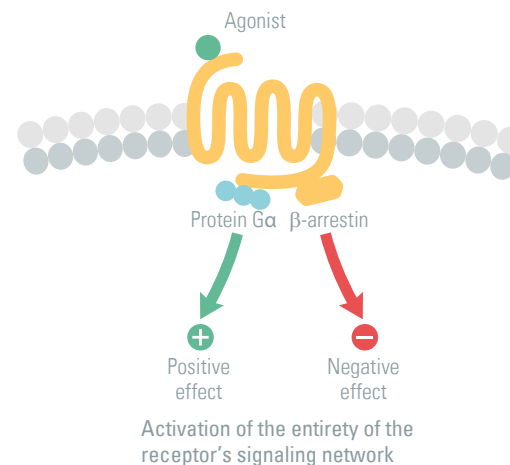
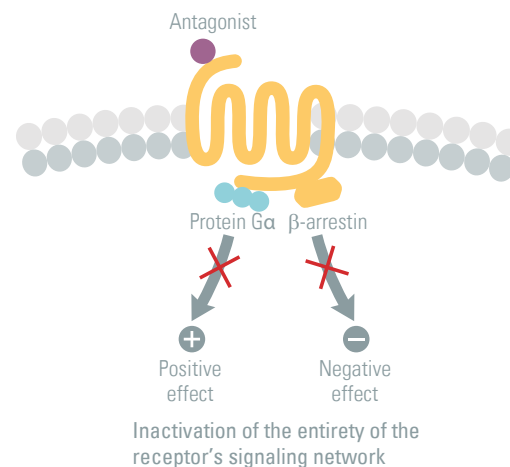


Biased agonism

Introduction

GPCR regulation has proven to be far more complex than previously thought by the scientific community. Evidence that GPCRs can couple to several effector pathways, and the existence of biased agonists able to activate them differentially, have introduced a new level of opportunity in GPCR drug research.

“Classical” agonists and antagonists respectively turn on, or off, the entirety of a receptor’s signaling network.

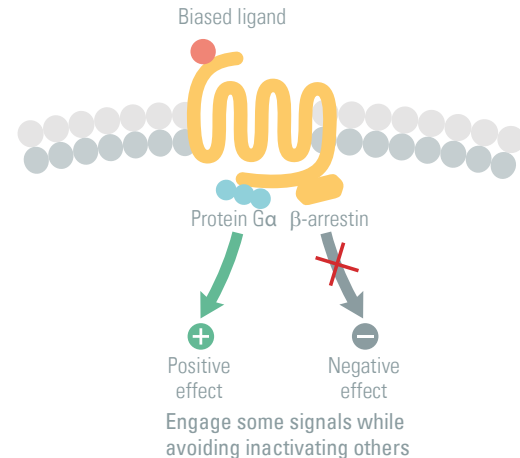


Biased ligands introduce a **new way to modulate GPCR function** in a finer manner and to separate therapeutic aspects from side-effects.

Nonetheless, it is still a challenge to identify and qualify ligands that activate only the pathways associated with the requested therapeutic benefit, and reliable tools are required. One interesting method for dissecting GPCR signal transduction concerns the Gαq signaling pathway.

IP-One offers significant advantages over traditional calcium assays including:

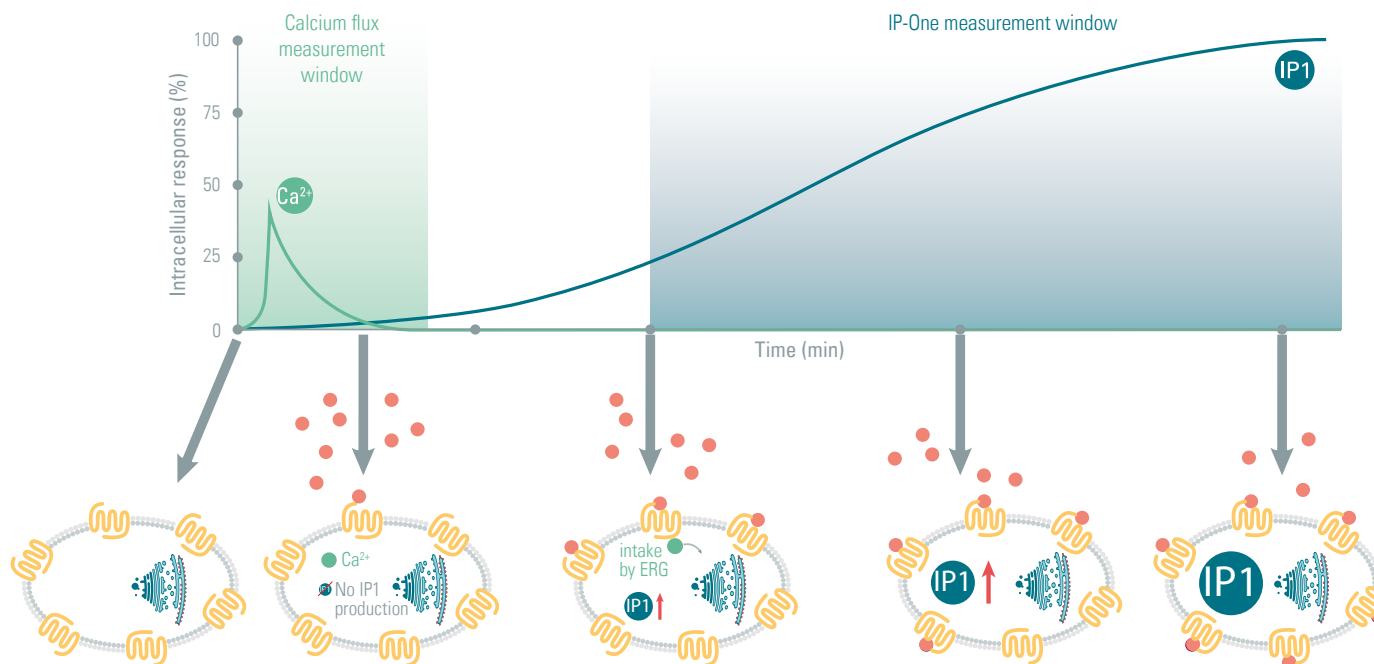
- IP-One is a «true equilibrium» assay
- IP-One is specific to Gαq signaling
- IP-One simplifies bias calculation
- IP-One effectively measures a receptor-proximate signal
- IP-One a reliable tool used by leading scientists worldwide



IP-One is a “true equilibrium” assay

Calcium transient assays are not ideal for characterizing drug bias on Gq coupled receptors. Due to the rapid nature of the calcium signal, flux assays are unlikely to reflect a true equilibrium state, a concept known as hemi-equilibrium. The interpretation of results may be hindered because of the non-equilibrium state. What may appear as functional selectivity could in reality stem from a lack of pre-equilibration. Bias calculation may thus be erroneous in the end.

Built on an accumulation assay, IP-One **distinguishes slow-acting agonists from true-biased agonists easily**, delivering more accurate data.

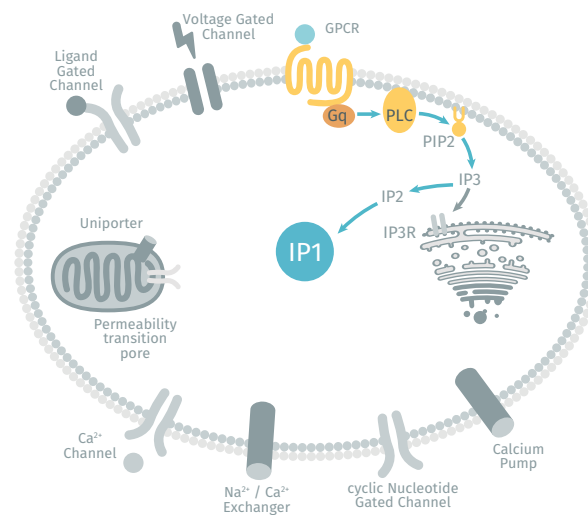
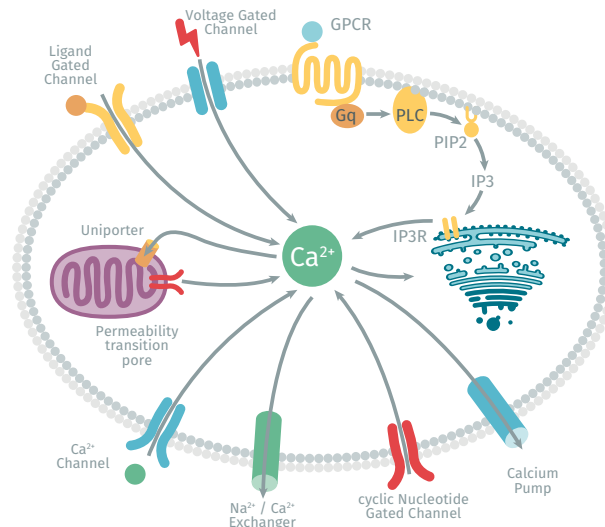


IP-One is **specific to Gq** signaling

Calcium responses are complex in nature. Calcium release is frequently triggered by non-G protein mechanisms, including Ca^{2+} permeable channels, calcium pumps, and calcium transporters.

IP-One ensures signal specificity. **IP-One only measures IP1**, an analyte that is solely produced by the action of the phospholipase enzyme (PLC) and mediated by GPCRs.

IP-One delivers with reliability and predictability, for results you can trust.



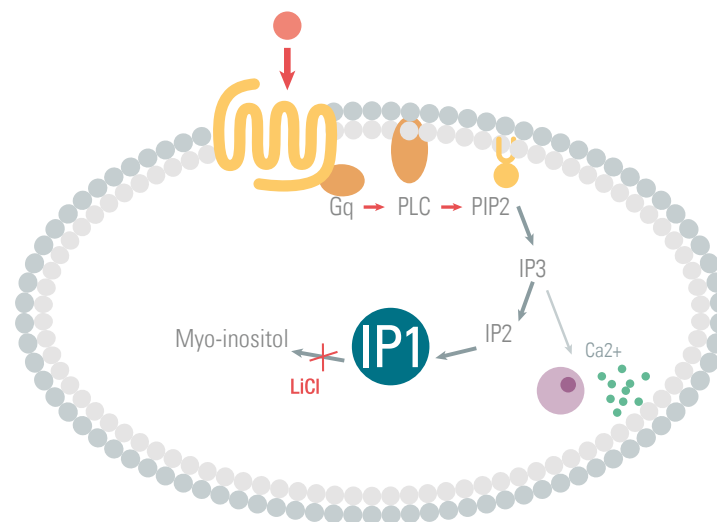
IP-One **simplifies bias** estimation and calculation

Signaling bias is a spatial concept. Bias may occur at different levels of the transduction cascade.

IP1 production occurs at a more proximal level of GPCR signal transduction than calcium release.

Bias estimation can benefit from the measurement of an event close to β -arrestin recruitment in the signal cascade.

Moreover, calcium assays deliver a fluorescence readout that cannot be converted easily in calcic concentration. The Cisbio IP-One assay rapidly delivers compound potency and efficacy in units of IP1 concentrations. The result is a simpler and more straightforward calculation of bias.



Literature

IP-One is a **reliable tool** commonly used by leading scientists and investigators.

A significant body of literature has been dedicated to IP-One.

Discover why leading academic authorities have relied on Cisbio's IP-One to advance their biased signaling research ^{[1][2][3]}.

Also read why private companies use IP-One to fuel their own drug development programs ^[4].

- [1] *Zimmerman, Brandon, et al.* «Differential β -arrestin-dependent conformational signaling and cellular responses revealed by angiotensin analogs.», *Science Signaling* 5.221 (2012): ra33-ra33.
- [2] *Kenakin, Terry, et al.* «A simple method for quantifying functional selectivity and agonist bias.», *ACS Chemical Neuroscience* 3.3 (2012): 193-203.
- [3] *Rajagopal, Sudarshan, et al.* «Quantifying ligand bias at seven-transmembrane receptors.», *Molecular Pharmacology* 80.3 (2011): 367-377.
- [4] *Kim, Ki-Seok, et al.* « β -arrestin-biased AT1R stimulation promotes cell survival during acute cardiac injury.», *American Journal of Physiology-Heart and Circulatory Physiology* 303.8 (2012): H1001-H1010.





Webinar

Biased ligands in Drug Discovery: the case of TRV027

A webinar discussing **Biased Ligands in Drug Discovery** featuring :

- **William Gowen-MacDonald** (R&D Scientist - Trevena),
- **Conrad Cowan** (Director of Biology - Trevena),
- **Terry Kenakin** (Professor - Dpt of Pharmacology UNC-CH),
- **Nicolas Pierre** (Corporate Product Manager - Cisbio)

is available to watch on demand.



Watch the full webinar
at [cisbio.com](https://www.cisbio.com)





Pharmacology

The success of your structure-activity relationships (SAR) campaigns is intimately dependent on the availability of pharmacologically accurate bioassays.

When these assays can reliably and precisely describe the drug receptor interactions involved, the chemical diversity of the leads can effectively be narrowed down and the best drug candidates selected with certainty.

Learn how IP-One delivers unbiased and accurate pharmacological profiles for results you can trust.



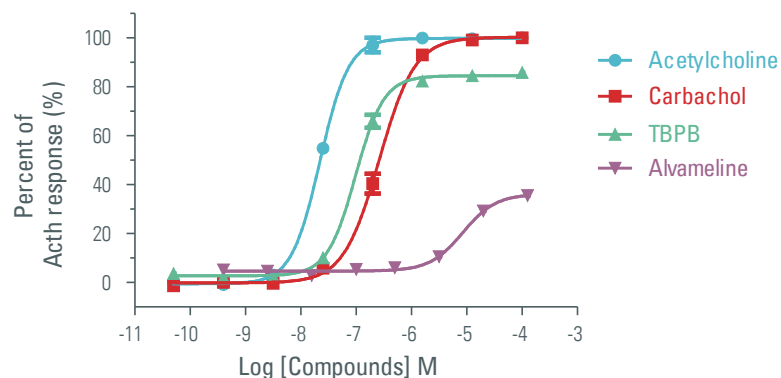


Compound characterization

Full and partial agonist

This graph shows a typical agonist mode screening assay using a CHO-M1 stable cell line model. Four well-known agonists, Acetylcholine, Carbachol, TBPB, and Alvimeline, have been characterized and ranked in terms of potency and efficiency.

The IP-One assay enables perfect characterizations and discrimination of full from partial agonist compounds.



Compounds	Class	Potency (nM)	Efficiency (%)
Acetylcholine	Full agonist	22	100
Carbachol	Full agonist	260	100
TBPB	Partial agonist	100	85
Alvimeline	Partial agonist	8500	35

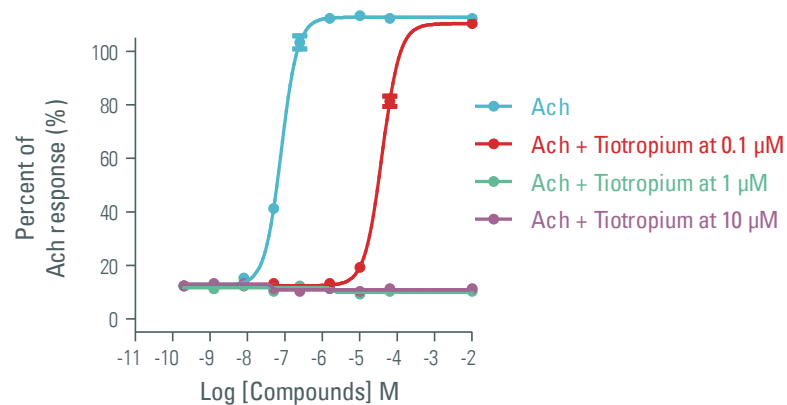


Antagonist

Tiotropium is a well-known insurmountable antagonist which binds on the orthosteric site of the M1 receptor.

Using a CHO-M1 stable cell line, the acetylcholine full agonist is incubated with or without different doses of tiotropium (0.1 μ M, 1 μ M, 10 μ M).

The results show a typical insurmountable antagonist pattern using the IP-One assay, as described in the literature*.



Compounds	EC50 (nM)
Acetylcholine	79
Acetylcholine + Tiotropium at 0.1 μ M	3971
Acetylcholine + Tiotropium at 1 μ M	Complete inhibition
Acetylcholine + Tiotropium at 10 μ M	Complete inhibition

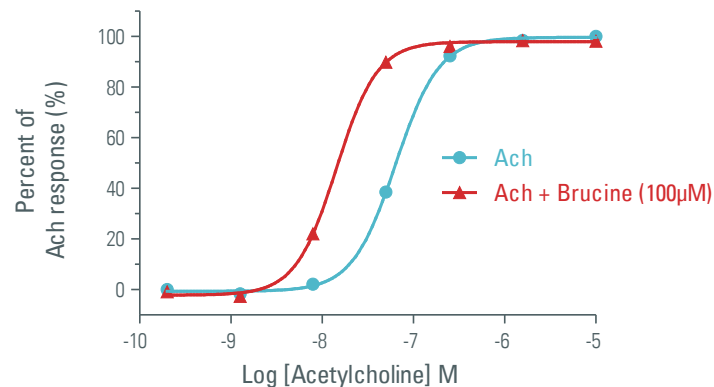
* Peter J. Barnes, MA, DM, DSc. The Pharmacological Properties of Tiotropium

Allosteric modulators

Brucine, a well-known PAM of the Muscarinic M1 receptor, was well characterized using the IP-One assay.

The CHO-M1 stable cells were treated with the acetylcholine full agonist alone or in addition to the Brucine positive allosteric modulator. Brucine acts by increasing the potency of the acetylcholine to the M1 receptor 4.6 fold at 100 μ M. and efficacy in units of IP1 concentrations.

The result is a simpler and more straightforward calculation of bias.



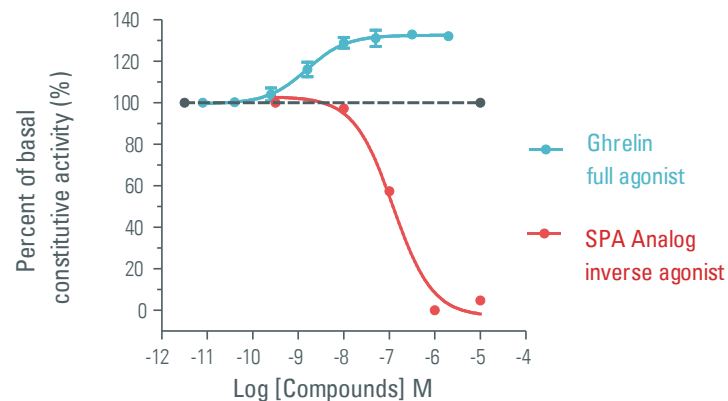
Compounds	Class	Potency of Ach (nM)	PAM effect
Acetylcholine	Full agonist	65	-
Acetylcholine + Brucine (100 μ M)	Full agonist + PAM	14	4.6 fold



Inverse agonist

The graph shows the profiling of a well-known inverse agonist of the GHSR1a receptor: SPA analog.

This compound inhibits the constitutive activity of the ghrelin receptor with high efficiency, reaching a level of no remaining constitutive activity (0%) after treatment.



Compounds	Class	Potency (nM)	% of basal constitutive activity
Acetylcholine	Full agonist	22	132
SPA analog	Inverse agonist	260	0

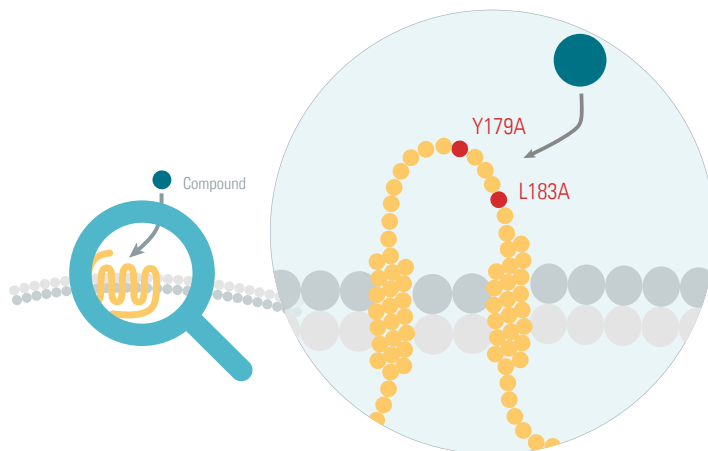
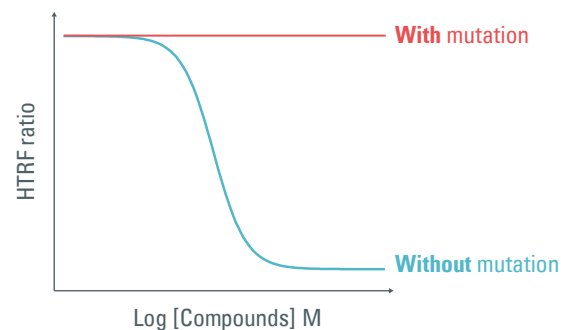


Alanine screens

Studying compound pharmacology on alanine mutants

IP-One is the preferred tool to study compound pharmacology on alanine mutants and will help you:

- Identify receptor selective ligand and understand the molecular basis of selectivity^[1]
- Identify orthosteric and allosteric ligand binding pockets
- Understand receptor determinants of potency and efficacy
- Obtain functional validation of computational modeling of ligand binding



[1] Abdul-Ridha, Alaa, et al. «Molecular determinants of allosteric modulation at the M1 muscarinic acetylcholine receptor.», Journal of Biological Chemistry 289.9 (2014): 6067-6079.





References

Literature

- *Abdul-Ridha, Alaa, et al.* «Molecular determinants of allosteric modulation at the M1 muscarinic acetylcholine receptor.», *Journal of Biological Chemistry* 289.9 (2014): 6067-6079.
- *Rovira, Xavier, et al.* «Overlapping binding sites drive allosteric agonism and positive cooperativity in type 4 metabotropic glutamate receptors.», *The FASEB Journal* 29.1 (2015): 116-130.
- *Siddiquee, K., et al.* «The apelin receptor inhibits the angiotensin II type 1 receptor via allosteric trans-inhibition.», *British journal of pharmacology* 168.5 (2013): 1104-1117.
- *Kuohung, Wendy, et al.* «A high-throughput small-molecule ligand screen targeted to agonists and antagonists of the G-protein-coupled receptor GPR54.», *Journal of biomolecular screening* 15.5 (2010): 508-517.
- *Matsufuji, Tetsuyoshi, et al.* «Discovery of novel chiral diazepines as bombesin receptor subtype-3 (BRS-3) agonists with low brain penetration.», *Bioorganic & medicinal chemistry letters* 24.3 (2014): 750-755.
- *Nordqvist, Anneli, et al.* «New hits as antagonists of GPR103 identified by HTS.», *ACS medicinal chemistry letters* 5.5 (2014): 527-532.
- *Yoshikawa, Keita, et al.* «Identification of alpha-substituted acylamines as novel, potent, and orally active mGluR5 negative allosteric modulators.», *Bioorganic & medicinal chemistry letters* 25.16 (2015): 3135-3141.





Webinar

Advancing GPCR drug discovery through allosteric modulation

A webinar reviewing experimental and analytical approaches for detecting and quantifying allosteric modulators featuring:

- **Meritxell Canals** (Ph.D. - Monash Institute of Pharm. Sci.),
- **J. Robert Lane** (Ph.D. - Monash Institute of Pharm. Sci.),
- **Terry Kenakin** (Professor - Dpt of Pharmacology UNC-CH),
- **Nicolas Pierre** (Corporate Product Manager - Cisbio)

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